



Published in final edited form as:

J Neurosurg Sci. 2014 September ; 58(3): 145–149. 以幹細胞及G-CSF治療腦急性損傷的神經發炎，可以合併治療之後的老化現象

Stem cells and G-CSF for treating neuroinflammation in traumatic brain injury: aging as a comorbidity factor

I. Dela Peña¹, P. R. Sanberg¹, S. Acosta¹, N. Tajiri¹, S.-Z. Lin², and C. V. Borlongan¹

¹Center of Excellence for Aging and Brain Repair, Department of Neurosurgery and Brain Repair, University of South Florida Morsani College of Medicine, Tampa, FL, USA

²Center for Neuropsychiatry, China Medical University and Hospital, No.2, Yude Road, Taichung, Taiwan, Republic of China

Abstract

Traumatic brain injury (TBI), often called the signature wound of Iraq and Afghanistan wars, is characterized by a progressive histopathology and long-lasting behavioral deficits. Treatment options for TBI are limited and patients are usually relegated to rehabilitation therapy and a handful of experimental treatments. Stem cell-based therapies offer alternative treatment regimens for TBI, and have been intended to target the delayed therapeutic window post-TBI, in order to promote “neuroregeneration,” in lieu of “neuroprotection” which can be accomplished during acute TBI phase. However, these interventions may require adjunctive pharmacological treatments especially when aging is considered as a comorbidity factor for post-TBI health outcomes. Here, we put forward the concept that a combination therapy of human umbilical cord blood cell (hUCB) and granulocyte-colony stimulating factor (G-CSF) attenuates neuroinflammation in TBI, in view of the safety and efficacy profiles of hUCB and G-CSF, their respective mechanisms of action, and efficacy of hUCB+G-CSF combination therapy in TBI animal models. Further investigations on the neuroinflammatory pathway as a key pathological hallmark in acute and chronic TBI and also as a major therapeutic target of hUCB+G-CSF are warranted in order to optimize the translation of this combination therapy in the clinic.

Keywords

Brain injuries; Stem cells; Aging

The wars in Iraq and Afghanistan have highlighted traumatic brain injury (TBI) as a significant unmet clinical need, characterized by high mortality and severe morbidity.^{1–4} Although traditionally considered as an acute event, TBI has now been recognized as being accompanied by chronic pathological symptoms, notably secondary cell death mediated by long-lasting neuroinflammation, and closely associated with life-long behavioral deficits.^{5, 6} To date, treatment for TBI is limited,⁷ with patients largely relegated to rehabilitation therapy.^{8–12}

In view of the rampant secondary cell death mediating the progression of TBI, novel treatments have targeted the delayed therapeutic time window post-TBI termed as “neuroregeneration” as opposed to the narrow “neuroprotection” window relegated to the acute TBI phase.^{4, 13} A major component of regenerative medicine is stem cell-based therapeutics which have been shown effective in animal models of neurological disorders, including TBI^{4, 14–17} and have reached limited trials in the clinic.¹⁸ Fetal stem cells, cancer-derived neuron-like cells, embryonic stem cells, induced pluripotent stem cells, and adult stem cells, such as umbilical cord blood, bone marrow stromal cells, amnion cells, have been examined for their safety, efficacy, and mechanisms of action for treating brain diseases.^{4, 14–18, 19–24} Our group, and several others, have assessed the clinical utility of human umbilical cord blood (hUCB)-derived cells in stroke, Parkinson’s disease, Huntington’s disease, cerebral palsy, and TBI.^{20, 25–28} Limited clinical trials of hUCB cells are being explored in cerebral palsy, inborn metabolic disorders, and stroke.^{26, 29–31}

In an effort to initiate clinical trials of hUCB cell therapy for TBI, translational research is necessary to determine the optimal transplant regimen and to provide a better understanding of the mode of action of stem cells in regenerating the injured brain. To this end, demonstrating a well-defined stem cell source is a basic translational gating item for quality assurance and quality control of graft origin, and also for validity and reproducibility of experimental outcomes. In the case of hUCB cells, there remains a paucity of research which identifies the appropriate cell population that is safe and effective for transplantation. There are reports implicating that the mononuclear fraction of hUCB exerts neuroprotective effects via multi-pronged neuroregenerative pathways including anti-inflammation and enhanced neurogenesis^{32–34}, while MSCs derived from hUCB have also been shown to promote functional benefits by increasing angiogenesis and vasculogenesis.^{35–37} In addition to identifying the optimal transplantable hUCB cell phenotype, low graft survival has been documented in the TBI brain, which may be due to the not-so-conducive host tissue likely created by the secondary neuroinflammatory response.^{38–40} While robust graft survival may not be necessary to induce behavioral recovery, the alternative mechanism of graft-induced by-stander effects still requires for the hostile microenvironment to be abrogated if improved clinical outcome is desired. Accordingly, the concept that stem cell therapy can be enhanced by rendering a receptive microenvironment (*i.e.*, less neuroinflammatory) appeals to advancing regenerative medicine for treating the injured brain.

Repurposing of old drugs poses as a logical approach when contemplating on testing adjunctive therapies for stem cell transplantation. An appealing drug candidate is the granulocyte-colony stimulating factor (G-CSF), an essential member of the hematopoietic growth factor family, which has received much attention over the last decade for its neuroprotective effects in animal models of stroke⁴¹ and Alzheimer’s disease⁴² via its action on reducing peripheral inflammation while stimulating neurogenesis.^{43, 44} Currently, a limited clinical trial is underway for testing G-CSF in TBI patients (personal communication with Dr. Juan Sanchez-Ramos, USF/VA). Notwithstanding inconsistent efficacy results with some studies showing significant functional recovery, while others reporting small incremental behavioral benefits G-CSF in TBI animals [e.g. 45,46], the translation of G-CSF

for TBI in the clinic is based on the drug's safety profiles for its indication in acute ischemic stroke and Alzheimer's disease.^{41, 42, 47}

Combining stem cell therapy and G-CSF for treating neuroinflammation in traumatic brain injury

The potential of combining stem cell transplantation with G-CSF to achieve improved therapeutic outcome in TBI has been recently examined.⁴⁸ Using a controlled cortical impact (CCI) model of moderate TBI in adult rats, we demonstrated that while monotherapy with hUCB or G-CSF promoted behavioral recovery, combined therapy of hUCB+G-CSF enhanced functional improvement compared to hUCB transplantation or G-CSF administration alone. In line with the concept of reducing neuroinflammation within the injured brain, co-administration with G-CSF might have produced a conducive microenvironment for the transplanted hUCB cells to integrate with the host tissue.⁴⁹ Immunohistochemical staining with OX-6, which labels MHCII+ activated microglia, revealed that hUCB+G-CSF reduced TBI-induced neuroinflammation in gray and white matter areas.⁴⁸ In addition, G-CSF might have directed the migration of endogenous stem cells mobilized from the bone marrow to the site of injury^{41-44, 46-49} and/or the grafted hUCB cells might have released growth factors secreted by hUCB grafts, and such combined regenerative pathways elicited a much more beneficial functional outcome than a single, stand-alone treatment.⁴⁸

Additive and synergistic effects likely mediated the improved outcomes produced by combined hUCB+G-CSF in TBI animals. Indeed, hUCB alone treatment reduces inflammation and promotes neurogenesis and angiogenesis,^{50, 51} while G-CSF alone enhances neurogenesis.⁴² Interestingly, the reduction in neuroinflammation exerted by hUCB+G-CSF therapy coincided with elevated neurogenesis in the dentate gyrus and subventricular zone of the hippocampus while increasing the survival of hippocampal neurons in TBI rats.⁴⁸ These findings suggest that hUCB+G-CSF synergistically diminished TBI-induced neuroinflammation and stimulated endogenous neurogenesis, while reducing cell death in the injured brain and promoting functional recovery in TBI animals.⁴⁸

As with any drug therapy, the blood-brain barrier penetrance and the ligand-receptor mechanism become translational gating items when advancing G-CSF therapy to the clinic. Of note, G-CSF is able to cross the blood-brain barrier and reach neurons and glial cells through the G-CSF receptor,⁵² and confers inhibitory actions upon pro-inflammatory cytokines while upregulating neurogenesis.^{41, 44, 52-55} In combination with stem cell therapy, G-CSF maintains stemness and directs differentiation of hUCB cells.⁵⁶ In parallel, G-CSF directed migration of mobilized bone marrow cells may interact with the transplanted hUCB cells involving a paracrine secretion of trophic factors, growth factors, chemokines and immune-modulating cytokines (also termed "bystander effects" of transplanted stem cells as noted above).^{57, 58} Such combination therapy of hUCB+G-CSF may thus entail a multipronged regenerative mechanism involving a receptor-mediated transport machinery with paracrine action in providing improved therapeutic outcome for TBI.

Aging exacerbates TBI-induced neuroinflammation: potential for cell therapy and G-CSF

Although the current theme of TBI has focused on our returning soldiers from the wars in the Iraq and Afghanistan, the aging population represents an equally large number of patients who suffer from TBI.^{59, 60} Thus, studies designed to incorporate aging as a comorbidity factor in TBI may provide a better understanding of the disease pathology, as well as the treatment regimen for this key patient population. In a recent study, mild TBI was performed in young (6-month-old) and aged (20-month-old) rats, and then transplanted intravenously (3 hours later) with 4×10^6 (6) human adipose-derived stem cells (Tx), conditioned media (CM), or vehicle (unconditioned media).⁴ The results showed significant recovery of motor and cognitive functions in young, but not aged, Tx and CM groups. Imaging analyses of hADSCs deposition revealed robust migration of Tx in the young spleen, but poorly reached the aged spleen. Moreover, while significant decrements in cortical damage and hippocampal cell loss were seen in both Tx and CM groups in young rats, only partial neuroprotection was observed in the aged rats and mainly in the Tx group but not the CM group. These findings suggest that while cell therapy appears effective for TBI, the reduced efficacy in aged rats, likely due to defective migration of the cells to the spleen, warrants optimization studies to improve the outcome of this treatment in the aging population. These suboptimal therapeutic effects solicit a similar combination therapy to supplement the functional benefits of stem cells transplantation. Because the spleen is a major source of systemic inflammation associated with neurological disorders, such as stroke and TBI,^{61–63} and is known to participate in the neuroprotective effects of cell therapy in TBI,^{64, 65} a drug that targets spleen-mediated inflammation is a logical candidate for such combination therapy with stem cells. As noted above, G-CSF stands as a robust neuroprotective agent, and additionally with highly potent anti-inflammatory effects.^{66–69} Recognizing that the aging brain when exposed to TBI may present with widespread inflammation, treatment with G-CSF may not only reduce the systemic inflammation, but may also harness a conducive host-brain microenvironment allowing the transplanted stem cells to exert their maximal therapeutic benefits.

Future directions

A progressive histopathology and long-lasting behavioral deficits characterize TBI. Rehabilitation therapy, and a handful of experimental treatments, remains the often utilized option for treating TBI patients.⁷⁰ The advent stem cell-based therapies offers alternative treatment regimens for TBI, but may require adjunctive pharmacological treatments, especially with the recognition that aging exacerbates TBI pathology rendering the brain hostile to stem cell survival and to the functional effects of transplanted stem cells. The combination therapy of hUCB+G-CSF appears beneficial in TBI animal models. The safety and efficacy profiles of hUCB and G-CSF, and their respective mechanisms of action, point to their entry into the clinic. Further investigations on the neuroinflammatory pathway as being a key TBI acute and chronic pathological hallmark, but also as a major therapeutic target of hUCB+G-CSF should optimize the application of this combination therapy for TBI.

Acknowledgments

Funding.—This study was funded by the Department of Defense W81XWH-11-1-0634, the University of South Florida Signature Interdisciplinary Program in Neuroscience funds, the University of South Florida and Veterans Administration Reintegration Funds, and the University of South Florida Neuroscience Collaborative Program. P. R. Sanberg and C. V. Borlongan have patents and patent applications on stem cell therapy and serve as founder and consultant, respectively, of Saneron CCEL Therapeutics, Inc. C. V. Borlongan is funded by the National Institutes of Health 1R01NS071956-01A1, and James and Esther King Biomedical Research Foundation 1KG01-33966. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

References

1. Acosta SA, Tajiri N, Shinozuka K, Ishikawa H, Grimmig B, Diamond D, Sanberg PR, et al. Long-term upregulation of inflammation and suppression of cell proliferation in the brain of adult rats exposed to traumatic brain injury using the controlled cortical impact model. *Plos One*. 2013; 8:e53376. [PubMed: 23301065]
2. Fabrizio KS, Keltner NL. Traumatic brain injury in operation enduring freedom/operation Iraqi freedom: a primer. *Nurs Clin North Am*. 2010; 45:569–580. [PubMed: 20971337]
3. Faul, M.; Xu, L.; Wald, MM.; Coronado, VG. Traumatic brain injury in the United States: Emergency department visits, hospitalizations and deaths. Atlanta (GA): Centers for Disease Control and Prevention, National Center for Injury Prevention and Control; 2010.
4. Tajiri N, Acosta SA, Shahaduzzaman MD, Ishikawa H, Shinozuka K, Pabon M, et al. Intravenous transplants of human adipose-derived stem cell protect the brain from traumatic brain injury-induced neurodegeneration and motor and cognitive and cognitive impairments: cell graft biodistribution and soluble factors in young and aged rats. *J Neurosci*. 2014; 34:313–316. [PubMed: 24381292]
5. Azouvi P, Vallat-Azouvi C, Belmont A. Cognitive deficits after traumatic coma. *Prog Brain Res*. 2009; 177:89–110. [PubMed: 19818897]
6. Wong D, Dahm J, Ponsford J. Factor structure of the depression anxiety stress scales in individuals with traumatic brain injury. *Brain Inj*. 2013; 27:1377–1382. [PubMed: 23924030]
7. Kaneko Y, Tajiri N, Yu S, Hayashi T, Stahl CE, Bae E, et al. Nestin overexpression precedes caspase-3 upregulation in rats exposed to controlled cortical impact traumatic brain injury. *Cell Med*. 2012; 4:55–63. [PubMed: 23101029]
8. Brasure M, Lamberty GJ, Sayer NA, Nelson NW, Macdonald R, Ouellette J, et al. Participation after multidisciplinary rehabilitation for moderate to severe traumatic brain injury in adults: a systematic review. *Arch Phys Med Rehabil*. 2013; 94:1398–1420. [PubMed: 23348125]
9. Giustini A, Pistarini C, Pisoni C. Traumatic and nontraumatic brain injury. *Handb Clin Neurol*. 2013; 110:401–409. [PubMed: 23312659]
10. Lu J, Gary KW, Neimeier JP, Ward J, Lapane KL. Randomized controlled trials in adult traumatic brain injury. *Brain Inj*. 2012; 13–14:1523–1548.
11. Rolan T. Traumatic brain injury? What do we know? *J Spec Oper Med*. 2013; 13:45–50. [PubMed: 24048989]
12. Twamley EW, Jak AJ, Dellis DC, Bondi MW, Lohr JB. Cognitive symptom management and rehabilitation therapy (CogSMART) for veterans with traumatic brain injury: pilot randomized controlled trial. *J Rehab Res Dev*. 2014; 51:59–70.
13. Mueller BK, Mueller R, Schoemaker H. Stimulating neuroregeneration as a therapeutic drug approach for traumatic brain injury. *Br J Pharmacol*. 2009; 157:675–685. [PubMed: 19422372]
14. Lindvall O, Brundin P, Widner H, Rehnström S, Gustavii B, Frackowiak R, et al. Grafts of fetal dopamine neurons survive and improve motor function in Parkinson's disease. *Science*. 1990; 247:574–577. [PubMed: 2105529]
15. Liu YP, Lang BT, Baskaya MK, Dempsey RJ, Vemuganti R. The potential of neural stem cells to repair stroke-induced brain damage. *Acta Neurophatol*. 2009; 117:469–480.

16. Mahmood A, Lu D, Lu M, Chopp M. Treatment of traumatic brain injury in adult rats with intravenous administration of human bone marrow stromal cells. *Neurosurgery*. 2003; 53:697–702. discussion 702–3. [PubMed: 12943585]
17. Yang M, Donaldson AE, Jiang Y, Iacovitti L. Factors influencing the differentiation of dopaminergic traits in transplanted neural stem cells. *Cell Mol Neurobiol*. 2003; 23:851–864. [PubMed: 14514036]
18. Huang HY, Chen L, Sanberg PR. Clinical achievements, obstacles, falsehoods, and future directions of cell-based neurorestoration. *Cell Transplant*. 2012; 21:S3–S11. [PubMed: 22507675]
19. Chen SJ, Chang CM, Tsai SK, Chang YL, Chou SJ, Huang SS, Tai LK, et al. Functional improvement of focal cerebral ischemia injury by subdural transplantation of induced pluripotent stem cells with fibrin glue. *Stem Cells Dev*. 2010; 19:1757–1767. [PubMed: 20192839]
20. Newman MB, Davis CD, Kuzmin-Nichols N, Sanberg PR. Human umbilical cord blood (HUCB) cells for central nervous system repair. *Neurotox Res*. 2003; 5:355–368. [PubMed: 14715454]
21. Newman MB, Misiuta I, Willing AE, Zigova T, Karl RC, Borlongan CV, et al. Tumorigenicity issues of embryonic carcinoma-derived stem cells: relevance to surgical trials using NT2 and hNT neural cells. *Stem Cells Dev*. 2005; 14:29–43. [PubMed: 15725742]
22. Sanberg PR, Eve DJ, Metcalf C, Borlongan CV. Advantages and challenges of alternative sources of adult-derived stem cells for brain repair in stroke. *Prog Brain Res*. 2012; 201:99–117. [PubMed: 23186712]
23. Sanberg PR, Eve DJ, Cruz LE, Borlongan CV. Neurological disorders and the potential role for stem cells as a therapy. *Br Med Bull*. 2012; 101:163–181. [PubMed: 22357552]
24. Sandoe J, Eggan K. Opportunities and challenges of pluripotent stem cell neurodegenerative disease models. *Nat Neurosci*. 2013; 16:780–789. [PubMed: 23799470]
25. Herranz AS, Gonzalo-Gobernado R, Reimers D, Asensio MJ, Rodriguez-Serrano M, Bazan E. Applications of human umbilical cord blood cells in central nervous system regeneration. *Curr Stem Cell Res Ther*. 2010; 5:17–22. [PubMed: 19807661]
26. Ilic D, Miere C, Lazic E. Umbilical cord blood cells: clinical trials in non-hematological disorders. *Brit Med Bull*. 2012; 102:43–57. [PubMed: 22544780]
27. Ou Y, Yu S, Kaneko Y, Tajiri N, Bae EC, Chheda SH, Stahl CE. Intravenous infusion of GDNF gene-modified human umbilical cord blood CD34+ cells protects against cerebral ischemic injury in spontaneously hypertensive rats. *Brain Res*. 2010; 1366:217–225. [PubMed: 20888805]
28. Garbozuva-Davis S, Willing AE, Saporta S, Bickford PC, Gemma C, Chen N, et al. Novel cell therapy approaches for brain repair. *Prog Brain Res*. 2006; 157:207–222. [PubMed: 17046673]
29. Copeland N, Harris D, Gaballa MA. Human umbilical cord blood stem cells, myocardial infarction and stroke. *Clin Med*. 2009; 9:342–345. [PubMed: 19728507]
30. Sanberg PR, Willing AE, Garbozuva-Davis S, Saporta S, Liu G, Sanberg CD, et al. Umbilical cord blood-derived stem cells and brain repair. *Ann N Y Acad Sci*. 2005; 1049:67–83. [PubMed: 15965108]
31. Verina T, Fatemi A, Johnston MV, Comi AM. Pluripotent possibilities: human umbilical cord blood cell treatment after neonatal brain injury. *Pediatr Neurol*. 2013; 48:346–354. [PubMed: 23583051]
32. Boltze J, Reich DM, Hau S, Reymann KG, Strassburger M, Lobsien D, Wagner DC, et al. Assessment of neuroprotective effects of human umbilical cord blood mononuclear cell subpopulations in vitro and in vivo. *Cell Transplant*. 2012; 21:723–737. [PubMed: 21929866]
33. Henning RJ, Shariff M, Eadula U, Alvarado F, Vasko M, Sanberg PR, Sanberg CD, et al. Human cord blood mononuclear cells decrease cytokines and inflammatory cells in acute myocardial infarction. *Stem Cells Dev*. 2008; 17:1207–1219. [PubMed: 18393684]
34. Pimentel-Coelho PM, Rosado-de Castro PH, Barbosa da Fonseca LM, Otero RM. Umbilical cord blood mononuclear cell transplantation for neonatal hypoxic-ischemic encephalopathy. *Pediatr Res*. 2012; 71:464–473. [PubMed: 22430382]
35. Acosta SA, Franzese N, Staples M, Weinbren NL, Babilonia M, Patel J, et al. Human umbilical cord for transplantation therapy in myocardial infarction. *J Stem Cell Res Ther*. 2013; 1(Suppl 4):S4-005. [PubMed: 24307973]

36. Lee EJ, Choi EK, Kang SK, Kim GH, Park JY, Kang HJ, et al. N-cadherin determines individual variations in the therapeutic efficacy of human umbilical cord blood-derived mesenchymal stem cells in a rat model of myocardial infarction. *Mol Ther.* 2012; 20:155–167. [PubMed: 22068423]
37. Li T, Ma Q, Ning M, Zhao Y, Hou Y. Cotransplantation of human umbilical cord-derived mesenchymal stem cells and umbilical cord blood-derived CD34⁺ cells in a rabbit model of myocardial infarction. *Mol Cell Biochem.* 2014; 387:91–100. [PubMed: 24166198]
38. Kumar A, Loane DJ. Neuroinflammation after traumatic brain injury: opportunities for therapeutic intervention. *Brain Behav Immun.* 2012; 26(8):1191–1201. [PubMed: 22728326]
39. Walker PA, Shah SK, Jimenez F, Aroom KR, Harting MT, Cox CS Jr. Bone marrow-derived stromal cell therapy for traumatic brain injury is neuroprotective via stimulation of non-neurologic organ systems. *Surgery.* 2012; 152:790–793. [PubMed: 22853856]
40. Woodlock T, Morganti-Kossmann MC. The role of markers of inflammation in traumatic brain injury. *Front Neurol.* 2013; 4:18. [PubMed: 23459929]
41. Shyu WC, Lin SZ, Yang HI, Yang HI, Tzeng YS, Pang CY, et al. Functional recovery of stroke rats induced by granulocyte colony-stimulating factor-stimulated stem cells. *Circulation.* 2004; 110:1847–1854. [PubMed: 15381647]
42. Sanchez-Ramos J, Song S, Sava V, Catlow B, Lin X, Mori T, Cao C, et al. Granulocyte colony stimulating factor decreases brain amyloid burden and reverses cognitive impairment in Alzheimer's mice. *Neuroscience.* 2009; 163:55–72. [PubMed: 19500657]
43. Schabitz WR, Kollmar R, Schwaninger M, Juettler E, Bardutzky J, Scholzke MN, et al. Neuroprotective effect of granulocyte colony-stimulating factor after focal cerebral ischemia. *Stroke.* 2003; 34:745–751. [PubMed: 12624302]
44. Schneider A, Kruger C, Steigleder T, Weber D, Pitzer C, Laage R, et al. The hematopoietic factor GCSF is a neuronal ligand that counteracts programmed cell death and drives neurogenesis. *J Clin Invest.* 2005; 115:2083–2098. [PubMed: 16007267]
45. Sakowitz OW, Schardt C, Neher M, Stover JF, Untenberg AW, Kiening KL. Granulocyte colony-stimulating factor does not affect contusion size, brain edema or cerebrospinal fluid glutamate concentrations in rats following controlled cortical impact. *Acta Neurochir Suppl.* 2006; 96:139–143. [PubMed: 16671442]
46. Yang DY, Chen YJ, Wang MF, Pan HC, Chen SY, Cheng FC. Granulocyte colony-stimulating factor enhances cellular proliferation and motor function recovery on rats subjected to traumatic brain injury. *Neurol Res.* 2010; 32:1041–1049. [PubMed: 20810026]
47. England TJ, Abaei M, Auer DP, Lowe J, Jones DR, Sare G, Walker M, et al. Granulocyte-colony stimulating factor for mobilizing bone marrow stem cells in subacute stroke: the stem cell trial of recovery enhancement after stroke 2 randomized controlled trial. *Stroke.* 2012; 43:405–411. [PubMed: 22198983]
48. Acosta S, Tajiri N, Shinozuka K, Ishikawa H, Sanberg P, Sanchez-Ramos J, et al. Combination therapy of human umbilical cord blood and granulocyte colony stimulating factor reduces histopathological and motor impairments in an experimental model of chronic traumatic brain injury. *PlosOne.* 2014; 9:e90953.
49. Willing AE, Vendrame M, Mallery J, Cassady CJ, Davis CD, Sanchez-Ramos J, et al. Mobilized peripheral blood cells administered intravenously produce functional recovery in stroke. *Cell Transplant.* 2003; 12:449–454. [PubMed: 12911133]
50. Iskander A, Knight RA, Zhang ZG, Ewing JR, Shankar A, Varma NR, et al. Intravenous administration of human umbilical cord blood-derived AC133⁺ endothelial progenitor cells in rat stroke model reduces infarct volume: magnetic resonance imaging and histological findings. *Stem Cells Transl Med.* 2013; 2:703–714. [PubMed: 23934909]
51. Shahaduzzaman M, Golden JE, Green S, Gronda AE, Adrien E, Ahmed A, et al. A single administration of human umbilical cord blood T cells produces long-lasting effects in the aging hippocampus. *Age (Dordr).* 2013; 35:2071–2087. [PubMed: 23263793]
52. Zhao LR, Piao CS, Murikinati SR, Gonzalez-Toledo ME. The role of stem cell factor and granulocyte-colony stimulating factor in treatment of stroke. *Recent Pat CNS Drug Discov.* 2013; 8:2–12. [PubMed: 23173646]

53. Hartung T. Anti-inflammatory effects of granulocyte colony-stimulating factor. *Curr Opin Hematol.* 1998;221–225. [PubMed: 9664164]
54. Morita Y, Takizawa S, Kamiguchi H, Uesugi T, Kawada H, Takagi S. Administration of hematopoietic cytokines increases the expression of anti-inflammatory cytokine (IL-10) mRNA in the subacute phase after stroke. *Neurosci Res.* 2007; 58:356–360. [PubMed: 17628734]
55. Toth ZE, Leker RR, Shahar T, Pastorino S, Szalavova I, Asemenev B, Key S, et al. The combination of granulocyte colony-stimulating factor and stem cell factor significantly increases the number of bone marrow-derived endothelial cells in brains of mice following cerebral ischemia. *Blood.* 2008; 111:5544–5552. [PubMed: 18268092]
56. Stachura DL, Svoboda O, Campbell CA, Espin-Palazon R, Lau RP, Zon LI, et al. The zebrafish granulocyte colony stimulating factors (Gcsfs): two paralogous cytokines and their roles in hematopoietic development and maintenance. *Blood.* 2013; 122:3918–3928. [PubMed: 24128862]
57. Borlongan CV. Bone marrow stem cell mobilization in stroke: a ‘bonehead’ may be good after all! *Leukemia.* 2011; 25:1674–1686. [PubMed: 21727900]
58. Yang M, Wei X, Li J, Heine LA, Rosenwasser R, Iacovitti L. Changes in host blood factors and brain glia accompanying the functional recovery after systemic administration of bone marrow stem cells in ischemic stroke rats. *Cell Transplant.* 2010; 19:1073–1084. [PubMed: 20412636]
59. Hawkins BE, Cowart JC, Parsley MA, Capra BA, Eidson KA, Hellmich HL, et al. Effects of trauma, hemorrhage and resuscitation in aged rats. *Brain Res.* 2013; 1496:28–35. [PubMed: 23274538]
60. Thurman DJ, Alverson C, Dunn KA, Guerrero J, Sniezek JE. Traumatic brain injury in the United States: A public health perspective. *J Head Trauma Rehabil.* 1999; 14:602–615. [PubMed: 10671706]
61. Li M, Li F, Luo C, Shan Y, Zhang L, Qian Z, et al. Immediate splenectomy decreases mortality and improves cognitive function of rats after severe traumatic brain injury. *J Trauma.* 2011;141–147. [PubMed: 21248654]
62. Rasouli J, Lekhrari R, Ozbalik M, Lalezari P, Casper D. Brain-spleen inflammatory coupling: a literature review. *Einstein J Biol Med.* 2011; 27:74–77. [PubMed: 22611344]
63. Vendrame M, Gemma C, Pennypacker KR, Bickford PC, Davis Sanberg C, Sanberg PR, et al. Cord blood rescues stroke-induced changes in splenocyte phenotype and function. *Exp Neurol.* 2006; 199:191–200. [PubMed: 16713598]
64. Walker PA, Shah SK, Harting MT, Cox CS Jr. Progenitor cell therapies for traumatic brain injury: barriers and opportunities in translation. *Dis Model Mech.* 2009; 2:23–28. [PubMed: 19132123]
65. Walker PA, Shah SK, Jimenez F, Gerber MH, Xue H, Cutrone R, Hamilton JA, et al. Intravenous multipotent adult progenitor cell therapy for traumatic brain injury: preserving the blood-brain barrier via an interaction with splenocytes. *Exp Neurol.* 2010; 225:341–352. [PubMed: 20637752]
66. Dietel B, Cicha I, Kallmünzer B, Tauchi M, Yilmaz A, Daniel WG, et al. Suppression of dendritic cell functions contributes to the anti-inflammatory action of granulocyte-colony stimulating factor in experimental stroke. *Exp Neurol.* 2012; 237:379–387. [PubMed: 22750328]
67. Kadota R, Koda M, Kawabe J, Hashimoto M, Nishio Y, Mannoji C, et al. Granulocyte colony-stimulating factor (G-CSF) protects oligodendrocyte and promotes hindlimb functional recovery after spinal cord injury in rats. *PLoS One.* 2012; 7:e50391. [PubMed: 23209732]
68. Solaroglu I, Cahill J, Tsubokawa T, Beskonakli E, Zhang JH. Granulocyte colony-stimulating factor protects the brain against experimental stroke via inhibition of apoptosis and inflammation. *Neurol Res.* 2009; 31:167–172. [PubMed: 19298757]
69. Sehara Y, Hayashi T, Deguchi K, Zhang H, Tsuchiya A, Yamashita T, et al. Decreased focal inflammatory response by G-CSF may improve stroke outcome after transient middle cerebral artery occlusion in rats. *J Neurosci Res.* 2007; 85:2167–2174. [PubMed: 17497673]
70. Lewis FD, Horn GJ. Traumatic brain injury: analysis of functional deficits and post-hospital rehabilitation outcomes. *J Spec Oper Med.* 2013; 13:56–61. [PubMed: 24048991]